

1 ***In vitro* morphogenesis, cryopreservation and induction of variability in bleeding heart**  
2 **(*Lamprocapnos spectabilis* (L.) Fukuhara): A review**

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8

9 **Abstract**

10 This review explores recent advances in the biotechnology of *Lamprocapnos spectabilis* (L.)  
11 Fukuhara (commonly known as bleeding heart), a valuable ornamental-medicinal perennial.  
12 The article covers *in vitro* morphogenesis, cryopreservation techniques, and methods for  
13 inducing variability. The establishment of *in vitro* cultures utilized Murashige and Skoog  
14 medium enriched with various auxins, cytokinins, gold nanoparticles, and plant extracts,  
15 under both fluorescent and wide-spectrum LED lighting. Axillary bud activation and indirect  
16 somatic embryogenesis were more efficient, particularly in the presence of kinetin and  
17 picloram, respectively, compared to adventitious shoot regeneration. Significant cultivar  
18 differences were observed, with 'Valentine' being the easiest and 'White Gold' the most  
19 challenging to culture *in vitro*. To mitigate stress caused by classical growth regulators,  
20 alternative substances such as nanoparticles and natural extracts were used. Gold  
21 nanoparticles enhanced shoot proliferation and plantlet quality, while coconut and rice  
22 extracts improved survival rates during acclimatization. Enhanced metabolite production was  
23 achieved using exogenous auxins and gold nanoparticles. Guaiacol peroxidase was identified  
24 as a sensitive oxidative stress marker, with glutathione reductase being the most stable under  
25 stress. Cryogenic techniques incorporating explant encapsulation, i.e. encapsulation-  
26 vitrification, showed high effectiveness and genetic stability of plants, with nanomaterials  
27 boosting effectiveness. Coconut extract also enhanced *post-thaw* shoot proliferation, while  
28 sesame extract served as a natural retardant for slow-growth cultures. Mutagenic effectiveness  
29 ranked as microwaves < nanoparticles < X-rays. Comprehensive genetic variability insights  
30 were provided by integrating multiple SPAR marker systems. This review underscores the  
31 promising biotechnological advancements for *L. spectabilis*, emphasizing the potential of *in*  
32 *vitro* techniques, innovative cryopreservation methods, and the application of nanoparticles  
33 and plant extracts to enhance micropropagation, genetic variability, and metabolite

34 production, thereby contributing to the conservation and commercial sustainability of this  
35 valuable ornamental-medicinal perennial.

36

37 **Keywords:** metabolism, nanoparticles, plant extracts, ornamental plants, stress reaction,  
38 tissue culture

39

## 40 **Introduction**

41 *Lamprocapnos spectabilis* (L.) Fukuhara is a herbaceous perennial native to Siberia, northern  
42 China, Korea, and Japan, where it has been known for at least 2000 years (Hodges 2012; Kim  
43 et al. 2018). Due to its unique flower shape, arranged in unilaterally pendulous racemes or  
44 spikes, this species is also known by the common names bleeding heart, lady in a bath, and  
45 lyre flower. In English literature, other terms such as fallopian buds and lady's locket also  
46 appear. According to various taxonomies, this species is placed in a separate, small botanical  
47 family Fumariaceae or the subfamily Fumarioideae belonging to Papaveraceae (Kamińska et  
48 al. 2005; Frey and Moretti 2019). In 1997, bleeding heart was reclassified (based on the  
49 morphology, as well as the data of nuclear ribosomal DNA internal transcribed spacer  
50 (nrDNA-ITS) and rps16 intron sequences) to the monotypic genus *Lamprocapnos* (Lidén et al.  
51 1997). Nevertheless, it is often referred to in literature and horticultural practice under its  
52 previous name *Dicentra spectabilis* (L.) Lem. (syn. *Fumaria spectabilis* L.) (Cho 2018; Igori  
53 et al. 2023). This cold-hardy species occurs in temperate climates, although it can also be  
54 found in south-central Alaskan home gardens (Robertson 2013). It is a long-day plant,  
55 flowering in late spring (April to June) - at the peak of the floristic season (Roberts et al.  
56 1995). The seeds of bleeding heart are black-colored and of spherical shape with white large  
57 elaiosome to be dispersed by ants (Kim et al. 2011). Mature plants produce fleshy tuberous  
58 roots (Kamińska et al. 2005). Due to its decorative leaves (varying shades of green or gold-  
59 yellow depending on the cultivar) and spectacular white, pink, or red flowers (Fig. 1),  
60 bleeding heart has been used in Europe and America for landscape architecture since the 19th  
61 century in plantings in parks, gardens, balconies, as a houseplant, and in floristry as a cut  
62 flower. The flowers are bisymmetric (as opposed to the actinomorphic flowers found in other  
63 Papaveraceae species), measuring 20–25 mm in width. They are flattened with a heart-shaped  
64 base and have two lanceolate, deciduous sepals that are 3–4 mm in length (Zhang and Zhao  
65 2018). Currently, methods for controlled cultivation of bleeding heart are known, allowing for  
66 year-round production *in vivo* (Hodges 2012). Due to the long vase life of cut flowers (lasting

67 from 8 to even 17 days), this species is particularly popular for Valentine's Day and Mother's  
68 Day (Roberts et al. 1995).

69 *L. spectabilis* may also find applications in medicine, pharmacology, and the cosmetic  
70 industry due to its high content of health-promoting isoquinoline alkaloids: protopine and  
71 sanguinarine (Och et al. 2017; Hyeon Kim et al. 2018; Adamski et al. 2020). The roots of  
72 bleeding heart are used in Asian folk medicine for treating ulcers and paralysis (Iwasa and  
73 Kim 1997). It has been also applied for the treatment of strokes, bruises, and blood circulation  
74 (Kim et al. 2017). The extract obtained from *L. spectabilis*, when applied to the skin even at  
75 low concentrations (0.1%), slows down UV-induced aging (Lee et al. 2004a). This species is  
76 also a source of antifungal and antibacterial substances, effective for example in combating  
77 methicillin-resistant *Staphylococcus aureus* F.J. Rosenbach strains (MRSA) (Ma et al. 2000).  
78 Studies conducted by McNulty et al. (2007) and Petruczynik et al. (2019) demonstrated the  
79 presence of substances with antidepressant properties in bleeding heart extracts, as well as  
80 biologically active lactones effective in eliminating human cancer cell lines (squamous cell  
81 carcinoma and adenocarcinoma). Therefore, it is worth paying more attention to this species  
82 and introducing it to *in vitro* conditions.

83 Tissue cultures can be used in plants for the following purposes: reproduction  
84 (micropropagation), obtaining secondary metabolites, storage and protection of genetic  
85 resources, improvement, and breeding (inducing variability). This technology is commonly  
86 used in various ornamental and medicinal plants, including bleeding heart (Rout and Jain  
87 2020). The objective of this review is to provide a comprehensive summary of the vegetative  
88 methods for *in vitro* propagation and long-term storage of *L. spectabilis* tissues in liquid  
89 nitrogen, while also exploring strategies to enhance genetic diversity within this species  
90 through both classical and modern biotechnological approaches.

91

## 92 **Development of a model system for vegetative propagation of bleeding heart *in vitro***

93 Bleeding heart is mainly propagated vegetatively through cuttings and division of fleshy  
94 roots, or less commonly through seeds (self-pollination can occur) (Sim et al. 2005; Hodges  
95 2012). Seed viability is low and they should be sown immediately after collection (Deno  
96 1993). Subsequently, the seeds must undergo a winter dormancy period to germinate in the  
97 spring of the following year. Thus, the entire developmental cycle of the plant takes nearly a  
98 year (Cho et al. 2020). Moreover, the generative propagation method does not guarantee the  
99 maintenance of maternal plant characteristics. Cutting and division *in vivo* ensure obtaining  
100 true-to-type clones and earlier flowering, however, the efficiency of these methods is limited

101 and dependent on climatic conditions. Additionally, the content of valuable secondary  
102 metabolites in plants cultivated *in vivo* is usually low and subject to significant fluctuations  
103 (Mohammad et al. 2014). *In vitro* technologies may provide a solution to these problems.

104         Until 2019, information on tissue cultures of bleeding heart was limited to the  
105 induction of indirect somatic embryogenesis from seeds or callus suspension culture in the  
106 presence of 2,4-dichlorophenoxyacetic acid (2,4-D) (Lee and Lee 2003; Lee et al. 2004b).  
107 However, somatic embryogenesis is a complex process associated with specific difficulties,  
108 such as asynchronous embryo development, lack of conversion into complete plants, and/or  
109 disruption of genetic stability in plants regenerated via callus. In studies by Lee and Lee  
110 (2003) and Lee et al. (2004b), 64% conversion of somatic embryos was achieved, of which  
111 only 46% survived acclimatization to greenhouse conditions. Moreover, suspension cultures  
112 are susceptible to contamination, and seed availability in bleeding heart is limited.  
113 Meristematic explants (apical and lateral buds) or non-meristematic explants (internodes,  
114 leaves, and their fragments) are much easier to obtain and allow the use of other  
115 micropropagation techniques besides the aforementioned somatic embryogenesis, i.e.,  
116 activation of lateral shoots and adventitious organogenesis (Fig. 2). These techniques allow  
117 for easy and quick production of a significant number of offspring plants. However, to fully  
118 utilize the potential of various micropropagation methods, it is necessary to conduct numerous  
119 experiments on different cultivars, considering factors such as type, age, and size of explant,  
120 type, and concentration of growth regulators, and their mutual interaction in the medium, as  
121 well as the influence of optical radiation (Moraes et al. 2021).

122

## 123 **Effect of growth regulators on the regeneration of meristematic explants**

### 124 ***Activation of axillary buds***

125 The morphogenetic response of meristematic explants (nodal segments) in bleeding heart was  
126 dependent on the genetic factor. For example, the proliferation and development of axillary  
127 shoots in the 'Gold Heart' cultivar were most effectively stimulated by kinetin (KIN), while  
128 for the 'White Gold' cultivar, MS medium (Murashige and Skoog 1962) with simultaneous  
129 addition of indole-3-butyric acid (IBA) and 6-benzyladenine (BA) proved to be the most  
130 optimal in terms of multiplication rate (9.8) and biometric parameters of shoots (Kulus 2020a,  
131 Kulus et al. 2021). These results are consistent with reports by Zagorskaya and Yegorova  
132 (2018) on the varied micropropagation efficiency of different plant cultivars. The relatively  
133 high dry matter content confirmed the high quality of the obtained microshoots without signs  
134 of hyperhydration, encountered in commercial laboratories (Vitroflora Ltd, Poland, personal

135 communication). Nevertheless, an additional subculture of shoots onto a medium containing  
136 only auxin proved necessary to induce rhizogenesis. 1-Naphthaleneacetic acid (NAA) was  
137 most effective in initiating this process, while Indole-3-acetic acid (IAA) stimulated root  
138 elongation. Despite the need to separate the multiplication and rooting stages, it was possible  
139 to obtain complete bleeding heart plants *in vitro* in a relatively short time (Kulus 2020a, Kulus  
140 et al. 2021). On the other hand, it was found that thidiazuron (TDZ), NAA, and picloram  
141 (PIC) inhibit caulogenesis in *L. spectabilis*, inducing intensive callus development instead  
142 (Kulus 2020a), which is consistent with reports on other plant species (Simon and Petrášek  
143 2011; Zhang et al. 2011; Kawochar et al. 2017).

#### 144 ***Somatic embryogenesis***

145 Indirect somatic embryogenesis was initiated after placing nodal explants on media containing  
146 PIC or NAA. According to Méndez-Hernández et al. (2019), auxins affect gene expression,  
147 including transcription factors (TF), which cause somatic cells to acquire embryogenic  
148 potential, which could explain the obtained results. In bleeding heart, somatic embryos were  
149 observed at all developmental stages described in scientific literature (Joshi and Kumar 2013).  
150 Additionally, groups of polyembryos and secondary adventitious embryos were formed. The  
151 highest number of somatic embryos per explant (12) was obtained on MS medium with 0.5  
152 mg·L<sup>-1</sup> PIC (Kulus 2020a). Considering that the explants used in the experiment were only 5-  
153 10 mm long, this method seems to be efficient for the micropropagation of bleeding heart or  
154 transgenesis research.

#### 155 ***Acclimatization***

156 Another crucial stage of micropropagation is plant acclimatization in the greenhouse (Fig.  
157 2E). The survival rate of bleeding heart microshoots after transfer to *ex vitro* conditions varied  
158 greatly, depending on the composition of the medium used for shoot multiplication (Kulus et  
159 al. 2021). A positive correlation was observed between the number of leaves on the shoot and  
160 the viability of acclimatized plants, which may be related to increased activity of the  
161 photosynthetic apparatus. A significant influence of the shoot multiplication medium  
162 composition on the quality of greenhouse-grown plants was also found. The presence of IBA  
163 in the multiplication medium was favorable with bleeding heart ‘White Gold’ in contrast to  
164 IAA-supplemented media. This effect, however, was not observed with the cultivar ‘Gold  
165 Heart’. Genetic analyses of RAPD (Randomly Amplified Polymorphic DNA (Williams et al.  
166 1990)) and SCoT (Start Codon Targeted Polymorphisms (Collard and Mackill 2009)) markers  
167 confirmed the stability of plants from experimental objects ensuring the highest multiplication

168 rate (Kulus et al. 2021). This fact is crucial for the possibility of using the developed protocols  
169 in commercial micropropagation of *L. spectabilis*.

170

### 171 **Effect of growth regulators on the regeneration of non-meristematic explants**

172 Due to the limited productivity of lateral meristems, Kulus and Tymoszuik (2020) focused on  
173 the possibility of using non-meristematic shoot fragments in *in vitro* systems of bleeding  
174 heart. Explants of the 'Alba' cultivar derived from whole leaves, petioles, and internodes were  
175 placed on a medium supplemented with BA, IAA, NAA, 2,4-D, or PIC in various  
176 concentrations and combinations.

### 177 ***Adventitious organogenesis***

178 The efficiency of callogenesis was independent of the explant type. However, the  
179 influence of explant type and medium composition on the dry and fresh mass of the obtained  
180 callus was confirmed. The results were surprising, as despite using 17 combinations of growth  
181 regulators, only 2.5% of explants regenerated adventitious shoots. Rhizogenesis was observed  
182 in 4.5% of explants (Kulus and Tymoszuik 2020). The obtained efficiency of adventitious  
183 organogenesis is lower than that described in other botanical families (Tymoszuik and Miler  
184 2019). This suggests that bleeding heart may be classified as a difficult species in  
185 micropropagation. Similar problems with stimulating *in vitro* regeneration have also been  
186 described in other members of the Papaveraceae (Park et al. 2004), which may be a  
187 characteristic feature of this family.

### 188 ***Somatic embryogenesis***

189 The efficiency of somatic embryogenesis in *L. spectabilis* was significantly higher,  
190 although dependent on the medium composition and explant type (somatic embryos were  
191 formed by 0 to 100% of explants). Embryogenic callus was most often formed on leaves and  
192 least often on internodes (Fig. 2D). The highest number of somatic embryos per explant (11.4)  
193 was formed on petioles on medium with 0.5 mg·L<sup>-1</sup> BA and 1.0 mg·L<sup>-1</sup> PIC (Kulus and  
194 Tymoszuik 2020). The auxin 2,4-D was also effective in inducing somatic embryogenesis.  
195 Joshi and Kumar (2013) report that synthetic auxins often trigger the embryogenic potential of  
196 cells and stimulate embryo proliferation at an early developmental stage while inhibiting the  
197 expression of genes responsible for their further maturation. These reports were partially  
198 reflected in studies on bleeding heart, as the majority of somatic embryos were indeed at an  
199 early developmental stage; nevertheless, germinating embryos with clearly visible embryonic  
200 roots were also observed (Kulus and Tymoszuik 2020).

201

202 **Multidirectional effects of plant extracts and nanoparticles (NPs) in micropropagation**  
203 **of bleeding heart**

204 Bleeding heart plants produced on media supplemented with traditional growth regulators  
205 often had noticeably paler leaf color compared to the untreated controls (Kulus et al. 2021). A  
206 probable explanation for this phenomenon is the adverse physiological changes caused by  
207 cytokinin BA, also described by other authors (Bidabadi and Jain 2020; Manokari et al. 2021).  
208 It was also found that the simultaneous addition of auxins and cytokinins to the medium (even  
209 at low concentrations) resulted in abundant polyphenol secretion (Fig. 2B), which inhibited  
210 plant growth (Kulus et al. 2021). Therefore, efforts were made to use substitutes for synthetic  
211 growth regulators in *L. spectabilis in vitro* cultures, i.e., plant extracts and nanoparticles.

212 Plant extracts can be a cheaper and more natural source of phytohormones, vitamins,  
213 nutrients (including sugars), phenols, and proteins beneficial for plants (Gnasekaran et al.  
214 2010, Markin et al. 2023). Some extracts may also contain natural growth retardants, which  
215 are an interesting alternative to osmotically active substances and synthetic compounds added  
216 to the medium for storing genetic resources under slow growth conditions. Over the years,  
217 extracts of various origins have been added to *in vitro* media mostly for orchid propagation  
218 (Molnár et al. 2011, Venkatachalam et al. 2015). In the experiment by Kulus and Miler  
219 (2021), the usefulness of traditional growth regulators and plant extracts obtained from  
220 coconut pulp and oat, rice, and sesame seeds in the activation and proliferation of lateral  
221 shoots of two *L. spectabilis* cultivars was compared. The latter three types of extracts were  
222 used for the first time in plant *in vitro* cultures. To learn more about their role in the  
223 morphogenetic response of explants, this study included an analysis of the chemical  
224 composition of the extracts and compared the costs of preparing the media (Kulus and Miler  
225 2021). The effect of the applied extracts on micropropagation efficiency usually depended on  
226 the cultivar, but also on the studied plant trait. Coconut extract stimulated shoot proliferation  
227 and increased the multiplication rate in the 'Gold Heart' cultivar. Rice extract, on the other  
228 hand, stimulated callogenesis in bleeding heart 'White Gold' and provided a nearly twofold  
229 higher multiplication rate than conventional growth regulators and, importantly, slowed down  
230 the plant aging process (Kulus and Miler 2021). A similar plant response to the addition of  
231 natural supplements to the medium was observed in *Pogostemon cablin* Benth. (Swamy et al.  
232 2014). An additional advantage of rice extract is its nearly four times lower cost of production  
233 compared to commercial auxins and cytokinins. This is of great importance, as the costs of  
234 purchasing reagents in laboratory plant production range from 20 to 40% of all financial  
235 outlays (Tomar et al. 2007; Chen 2016). Sesame extract, in turn, inhibited the development of

236 explants in both studied cultivars, probably due to the relatively high content of polyphenols  
237 (Kulus and Miler 2021). Therefore, it can be successfully used in the slow-growth culture of  
238 bleeding heart. An additional advantage of using natural plant products was the easier  
239 disposal of used medium, which after appropriate processing, can be used as fertilizer in  
240 further *ex vitro* cultivation.

241 A breakthrough in developing micropropagation protocols for *L. spectabilis* was the  
242 use of nanomaterials by Kulus et al. (2022). Nanoparticles are structures with dimensions  
243 from 1 to 100 nm. Compared to conventional materials, they are characterized by higher  
244 chemical reactivity and unique physical properties (Fayez et al. 2017). It is known that  
245 nanoparticles can interact directly with the cell, affecting plant growth and metabolic activity  
246 positively or negatively, depending on the parameters of the nanoparticles, i.e., their type,  
247 concentration, method of synthesis, diameter, shape, etc., as well as the properties of the plant  
248 material itself (genotype, organ, anatomical structure) (Sanzari et al. 2019). Currently, many  
249 types of nanoparticles are synthesized, among which silver nanoparticles (AgNPs) are most  
250 often used in plant research (Tymoszuk and Miler 2019). Overall, however, there are still  
251 many unknowns regarding the impact of nanoparticles on living organisms and the  
252 environment, especially on the possibility of using them in *in vitro* cultures. The addition of  
253 gold nanoparticles to the medium, regardless of their concentration (50 – 100 ppm), positively  
254 affected not only the proliferation of lateral shoots of bleeding heart ‘Valentine’ but also their  
255 further growth *in vitro* (Fig. 2C) compared with the plane MS medium (Fig. 2A). The  
256 multiplication rate in the presence of NPs reached a value close to 23, more than twice as high  
257 as in the case of classical growth regulators. No effect of nanoparticles on the relative water  
258 content in shoots or the efficiency of rhizogenesis in the rooting medium (100%) was found.  
259 Nanoparticles, however, stimulated elongation and "branching" of roots, which emphasizes  
260 their usefulness (Kulus et al. 2022).

## 261

### 262 **Effect of wide-spectrum light-emitting diodes on the *in vitro* morphogenesis of bleeding** 263 **heart**

264 Light is a crucial factor influencing plant growth and development (Paik and Huq 2019).  
265 Currently, increasing attention is being paid to the possibility of using modern light sources  
266 such as LEDs (Light Emitting Diodes) in micropropagation. Apart from economic and  
267 environmental aspects (longer diode lifespan, lower power consumption, and less heat  
268 emission compared to traditional fluorescent lamps), the advantage of using LEDs in plant  
269 cultivation is the ability to precisely control the optical spectrum. Previous plant studies have



270 mainly focused on red and blue light (Bello-Bello et al. 2017). However, the latest research  
271 indicates that plants require a richer spectrum for full functional efficiency.

272 In the article by Miler et al. (2019), the impact of optical spectrum composition on the  
273 effectiveness and economy of micropropagation of five ornamental plant species, including  
274 bleeding heart, was verified. For this purpose, wide-spectrum LEDs and standard fluorescent  
275 lamps were used. The results indicated that the selection of light conditions in the growth  
276 room/phytotron should be adapted to the individual needs of particular plant species, which is  
277 consistent with reports by Gupta and Jatothu (2013). LED modules with high content of red  
278 and far-red light, crucial for photosynthesis efficiency, are optimal for the *in vitro* growth and  
279 micropropagation of bleeding heart. Such a spectrum allows for obtaining a multiplication  
280 rate and plant biometric parameters comparable to those observed under fluorescent lamps,  
281 with simultaneously twice lower electricity consumption. Lamps with increased blue and  
282 green light content do cause a significant reduction in bleeding heart shoot length, but the  
283 electricity costs necessary to produce 1 million plants using these modules are also four times  
284 lower (Miler et al. 2019). This is particularly important from the producers' point of view, as  
285 electricity costs are among the highest in commercial *in vitro* laboratories (Chen 2016) and  
286 can constitute from 20% to even 60% of all micropropagation costs, depending on the  
287 geographical location of the laboratory (Tomar et al. 2007). Regardless of the light source  
288 used, it was not possible to induce simultaneous rooting of plants at the multiplication phase.  
289 For comparison, the closely related *Dicentra × hybrida* regenerates complete shoots and roots  
290 on MS medium without the addition of growth regulators and under classic fluorescent lamps,  
291 with 100% acclimatization success (Kulus 2021). This emphasizes the difficulty of cultivating  
292 *L. spectabilis* under *in vitro* conditions.

293

#### 294 **Development of cryopreservation protocols and their impact on the stability of *L.*** 295 ***spectabilis***

296 Biodiversity conservation is one of the most important challenges of the modern world.  
297 Although commercial cultivars of bleeding heart are popular in cultivation, the number of  
298 endemic populations of this species is small, and it may be classified as endangered in the  
299 future (Hammer et al. 2005). For short-term storage, propagation, and transport, artificial  
300 seeds can be useful (Gantait et al. 2015). However, cryopreservation, which involves storing  
301 tissues at the cryogenic temperature of liquid nitrogen (-196°C), is considered the most  
302 effective method for long-term biodiversity conservation (El Merzougui et al. 2023; Sochacki  
303 et al. 2024). This approach can also be useful for storing cell lines with a particularly high

304 capacity for producing valuable secondary metabolites. In the available literature before 2019,  
305 there was no information on the use of artificial seeds and cryogenic techniques for *L.*  
306 *spectabilis*. Maintaining viable biological material in liquid nitrogen is possible, provided that  
307 the tissues are properly prepared (El Merzougui et al. 2023). Cryopreservation is a complex  
308 process that requires careful optimization of several key stages. The choice of  
309 cryopreservation method (vitrification, droplet-vitrification, encapsulation-vitrification, or  
310 encapsulation-dehydration) depends on the species. It is then necessary to optimize the  
311 composition of the pre-culture medium, the first stage of the procedure, which aims to pre-  
312 harden the cells. In the next stage, it is crucial to select the appropriate plant vitrification  
313 solution (PVS), a mixture of dehydrating, protective, and stabilizing substances  
314 (cryoprotectants). The concentration of PVS and the treatment time must be considered. In the  
315 encapsulation-dehydration technique, it is necessary to optimize the drying time. The prepared  
316 plant material is placed in liquid nitrogen, but to regenerate a complete plant, the thawed  
317 explants must be placed on a growth medium with an appropriately selected composition  
318 (Popova et al. 2023).

319 Kulus (2020b) aimed to develop a simple cryopreservation procedure based on drying  
320 explants with sterile air, without the need for chemical cryoprotectants. However, it was  
321 found that the apical buds of *L. spectabilis* have limited tolerance to prolonged desiccation.  
322 Despite considering several experimental factors, the effectiveness of this technique was  
323 limited (survival rate of 31.3 – 36.4%). Interestingly, the explants that survived freezing  
324 produced longer shoots with significantly greater mass than untreated controls. The  
325 improvement in plant quality was also evident during the planimetric analysis of leaf size  
326 (Kulus 2020b). A similar positive effect of stress on plant development was described by  
327 Adamczuk et al. (2012) in the *in vitro* cultures of flax and oat. The observed beneficial effect  
328 of encapsulation on the growth and development of non-frozen explants highlights the  
329 relevance of using artificial seeds for storage, transport, and propagation of bleeding heart.

330 Given the limited effectiveness of the encapsulation-dehydration technique, three other  
331 cryopreservation methods were used: vitrification, droplet-vitrification, and encapsulation-  
332 vitrification (Kulus 2020c, d). The regenerated plant material was then subjected to detailed,  
333 multi-level analysis using various analytical methods. Comparative analyses indicated that  
334 vitrification solution 3 (PVS3) (Nishizawa et al. 1993) was the most effective in securing  
335 cryo-treated tissues of *L. spectabilis*, both in terms of explant survival and the quality of the  
336 plants obtained from them (Kulus 2020d). Furthermore, numerous experiments revealed that  
337 the tolerance of encapsulated bleeding heart explants to prolonged dehydration is significantly

338 higher than that of non-encapsulated apical buds, regardless of storage in liquid nitrogen  
339 (Kulus 2020c). The relevance of supplementing the alginate capsule with salts and vitamins  
340 from the MS medium and the impact of this treatment on internode elongation and leaf  
341 development was also highlighted (Kulus 2020d). The encapsulation-vitrification technique,  
342 based on shoot tip (2-4 mm) preculture on the MS medium with 1.0 mg·L<sup>-1</sup> KIN, 9% sucrose,  
343 and 10 μM abscisic acid (ABA), encapsulation in 3% calcium alginate, followed by 20 min  
344 exposure to loading solution (LS, 2.0 M glycerol and 0.4 M sucrose) and 150-min dehydration  
345 with PVS3, ensured the highest survival rate of biological material (over 73%), as well as the  
346 most intensive plant development after thawing explants, including shoot proliferation and  
347 root regeneration, which was not observed in the control object. The lowest cryopreservation  
348 efficiency was obtained with the vitrification technique (Kulus 2020c). Interestingly, the  
349 opposite results were reported in *Chrysanthemum × morifolium* /Ramat./ Hemsl. ‘Escort’  
350 (Halmagyi et al. 2004). Another advantage of the developed procedure was the stable viability  
351 of explants, which did not change in time. All viable apical buds resumed growth, which is  
352 not always evident in post-freezing plant culture (Osorio-Saenz et al. 2011).

353 Spectral analyses conducted in the next stage of the research helped to understand the  
354 impact of the cryopreservation procedure on the biochemical activity of plants (Kulus 2020c;  
355 Kulus and Tymoszuik 2021). The chlorophyll content was significantly lower in shoots  
356 obtained from cryopreserved meristems compared to the control, which is likely due to the  
357 high lability of these pigments described by Van Assche and Clijsters (1990). It was also  
358 found that in response to moderately strong stress, *L. spectabilis* plants exhibit increased  
359 anthocyanin production, which significantly decreases with prolonged exposure to the stressor  
360 (Kulus 2020c).

361 The ultimate indicator of the effectiveness of the cryopreservation procedure is the  
362 absence of genetic changes in the stored biological material (Adhikari et al. 2020). ISSR  
363 (Inter Sequence Simple Repeat (Zietkiewicz et al. 1994)), RAPD, and SCoT molecular  
364 analyses, optimized for this species by Kulus (2020b, c), demonstrated the effectiveness of  
365 encapsulation-based methods in securing genetic stability (100% DNA sequence homology  
366 with untreated control). Slight genetic variation was detected by RAPD and ISSR markers  
367 after applying other cryogenic techniques, as well as in the untreated control (Kulus 2020c).  
368 Similar variability induced by cryo-treatment was detected in *Hladnikia pastinacifolia* Rchb.  
369 (Ciringer et al. 2018) and several other plant species (Kulus and Mikuła 2016). Therefore, it  
370 can be concluded that this phenomenon is not extremely rare. Interestingly, polymorphic  
371 bands detected in the genotype obtained after cryopreservation were also identified in one

372 control plant (Kulus 2020c). This may indicate the presence of so-called hot spots in the  
373 genome of bleeding heart, which are particularly prone to mutations (Rogozin and Pavlov  
374 2003). The detected polymorphisms in the non-cryopreserved control object indicate that  
375 long-term storage of *L. spectabilis* genetic resources should be performed under cryogenic  
376 conditions.

377

### 378 **Application of plant extracts and gold nanoparticles in cryopreservation of *L. spectabilis*** 379 **shoot tips**

380 The effectiveness of cryogenic techniques can be modified using unconventional  
381 supplements, although knowledge in this area is limited. Kulus and Miler (2021) verified the  
382 usefulness of plant extracts in the cryopreservation of bleeding heart ‘Gold Heart’ and ‘White  
383 Gold’. For this purpose, coconut, oat, rice, or sesame extract was added to the preculture  
384 medium during the first stage of the encapsulation-vitrification cryopreservation procedure.  
385 The use of extracts did not improve the survival of shoot tips stored in liquid nitrogen, and oat  
386 and sesame extracts even lowered the value of this parameter (Kulus and Miler 2021). This  
387 may result from the diverse fatty acid profile in individual extracts. The saturation level of  
388 fatty acids affects their physicochemical properties, and thus also the uptake of nutrients and  
389 cryoprotectants from the medium by explants (Mei et al. 2015). Despite the lack of  
390 improvement in tissue survival, coconut extract stimulated more intensive proliferation and  
391 shoot development after thawing the explants, indicating its usefulness in further research.  
392 This can be explained by the fact that coconut extract is rich in endogenous cytokinins and  
393 simple sugars (participating, among others, in the formation of so-called biological glass in  
394 the vitrification process) (Yong et al. 2009). Despite their potentially valuable effect,  
395 optimizing the concentrations and doses of natural plant extracts in plant tissue culture  
396 presents significant challenges due to the variability in phytochemical composition and  
397 biological activity among different plant sources. Additionally, the effects of these extracts  
398 can be influenced by factors such as the specific plant species, developmental stage, and  
399 culture conditions, making it difficult to establish standardized protocols for effective  
400 application (Hamdeni et al. 2022).

401 The effectiveness of cryopreservation can be improved using nanoparticles. One of the  
402 unique properties of nanoparticles is their high thermal conductivity (Li et al. 2005), which is  
403 crucial for the success of cryopreservation at the stage of freezing and thawing tissues. In the  
404 study by Kulus and Tymoszuk (2021) and Kulus et al. (2024), shoot tips of bleeding heart  
405 ‘Gold Heart’ and ‘Valentine’ were subjected to cryopreservation using the encapsulation-

406 vitrification method. Gold (AuNPs), silver (AgNPs), or zinc oxide (ZnONPs) nanoparticles  
407 were added at various concentrations to the preculture medium, or to the protective alginate  
408 coating, or to the *post-freezing* growth medium. To exclude the possibility of undesirable  
409 interactions that could disturb the unambiguity of the effect of the studied factor, the research  
410 included a cryotreated positive control (treated only with nanoparticle synthesis stabilizer), a  
411 cryotreated negative control (not treated with either stabilizer or AuNPs), and a standard not  
412 subjected to cryopreservation. Plants recovered from explants stored in liquid nitrogen were  
413 subjected to extensive stability analysis.

414 The addition of AuNPs or ZnONPs (13 nm in diameter) or AgNPs (at 6 nm) at low  
415 concentrations to the alginate bead significantly increased the effectiveness of  
416 cryopreservation (over 20% increase in explant survival compared to the untreated control),  
417 without affecting the genetic stability of plants (Kulus and TymoszuK 2021; Kulus et al.  
418 2024). This was probably due to the accelerated rate of cooling and thawing of tissues  
419 provided by nanometals (Vanzha et al. 2016). Supplementation of preculture medium with  
420 NPs did not cause an improvement in explants viability, although the produced shoots were  
421 significantly longer (Kulus et al. 2024). Conversely, the addition of nanoparticles to the *post-*  
422 *freezing* recovery medium had a negative impact on the survival of bleeding heart shoot tips.  
423 The identification of structural-metabolic changes in the *in vitro* cultures suggests that this  
424 effect resulted from the disruption of cell membrane integrity and the induction of oxidative  
425 stress in cells by aggregating nanoparticles. The negative impact of AuNPs intensified with  
426 the increase in their concentration in the medium. Additionally, it was found that the change  
427 in biochemical activity of cells and the morphogenetic response of explants (especially the  
428 rooting efficiency of plants and root parameters) depended on the moment of nanoparticle  
429 application (before or after liquid nitrogen treatment) (Kulus and TymoszuK 2021). Generally,  
430 however, the research hypothesis on the usefulness of nanoparticles in cryopreservation was  
431 confirmed, which has great significance and can be used by a wide range of scientists. A  
432 comparison of the efficiency of various cryopreservation protocols is given in Figure 3. The  
433 obtained results also emphasize the validity of using nanoparticles in the production of  
434 artificial seeds. Interestingly, it was found that under suboptimal conditions, nanoparticles  
435 exhibited genotoxic properties (Kulus and TymoszuK 2021). This suggests that at sufficiently  
436 high concentrations, NPs can be used as a chemical mutagen useful in bleeding heart breeding  
437 programs.

438

439 **Enzymatic and non-enzymatic cellular responses to *in vitro* culture conditions**

440 The metabolic activity of biological material is among the most critical factors determining  
441 the efficiency of *in vitro* systems, especially with medicinal crops. *In vitro* cultivation of *L.*  
442 *spectabilis* could have significant potential for the overproduction of valuable medicinal  
443 metabolites, particularly through the application of various elicitors that enhance biosynthetic  
444 pathways (Fazili et al. 2022). By optimizing culture conditions and utilizing substances like  
445 auxins, cytokinins, or nanoparticles, researchers could successfully increase the concentration  
446 of key compounds, such as tannins or antioxidants (Thakur et al. 2019). Moreover, the  
447 metabolic activity of plants, and their capability to produce primary and secondary  
448 metabolites, affects micropropagation and cryopreservation efficiencies.

449 Chlorophyll content, as the main component of chloroplasts, influences the efficiency  
450 of photosynthesis and the physiological activity of plants, affecting their growth *ex vitro*.  
451 However, stress conditions can lead to a reduction in this pigment, directly associated with  
452 decreased photosynthesis rates, ultimately inhibiting plant growth and development (Sherin et  
453 al. 2022). Flavonoids and polyphenols are antioxidants produced by plants mainly to protect  
454 them against stress and participate in cell detoxification by acting as metal chelators and  
455 neutralizing reactive oxygen species (ROS) (Thiruvengadam et al. 2015). Therefore, it is  
456 essential to monitor the physiological state and quality of plants both during *in vitro*  
457 cultivation and after transfer to greenhouse conditions (Halder et al. 2019).

458 To better understand the physiological responses of *L. spectabilis*, shoots and callus  
459 obtained *in vitro* were subjected to biochemical analysis for the content of pigments crucial  
460 for photosynthesis and non-enzymatic antioxidants (Kulus 2020a; Kulus et al. 2020; Kulus  
461 and Tymoszuik 2020). Additionally, Kulus and Tymoszuik (2021) and Kulus et al. (2022)  
462 determined the activity of selected antioxidant enzymes in *L. spectabilis* plants. This provided  
463 valuable information on the tolerance levels of *L. spectabilis* to specific physical and chemical  
464 factors and the mechanisms activated in response to severe abiotic stress.

#### 465 ***Effect of plant growth regulators and nanoparticles on the content of metabolites in shoots***

466 The study demonstrated a significant impact of *in vitro* culture conditions on the metabolite  
467 content in *L. spectabilis* shoots, with this effect depending on the cultivar and the treatment  
468 method. Auxin IAA stimulated the biosynthesis of chlorophyll and carotenoids in the 'Gold  
469 Heart' cultivar, while none of the classic growth regulators increased the content of these  
470 pigments in 'White Gold' plants (Kulus 2020a), highlighting the difficulty of growing this  
471 cultivar *in vitro*. In some experimental objects (especially in the presence of BA and KIN), a  
472 decrease in photosynthetic pigment content in shoots was observed, particularly if callus  
473 formed at their base (Kulus et al. 2020). This is surprising, as cytokinins typically participate

474 in plastid development and differentiation, and pigment biosynthesis (Dobránszki and  
475 Mandler-Drienyovszki 2014). The treatment method significantly influenced the anthocyanin  
476 and total polyphenol content in shoots, while it had a lesser effect on other flavonoid  
477 compounds.

478 On the other hand, it was found that dehydration (PVS treatment) and  
479 cryopreservation (regardless of the technique) resulted in a decreased synthesis of  
480 chlorophylls, confirming the high lability of this pigment group. Production of anthocyanins  
481 was more varied and dependent on the protocol used (Kulus 2020c).

482 Nanoparticles have been shown to significantly influence the production of  
483 metabolites in *L. spectabilis*. Gold nanoparticles were found to act as elicitors of tannin  
484 biosynthesis in *L. spectabilis* shoot cultures (Kulus et al. 2022). Moreover, Kulus et al. (2024)  
485 reported that the incorporation of AuNPs, AgNPs, and ZnONPs into the preculture medium  
486 during the encapsulation-vitrification cryopreservation protocol enhanced the biosynthesis of  
487 primary and secondary metabolites, including pigments such as chlorophyll and carotenoids,  
488 in a cultivar-specific manner, with notable increases observed in the 'Gold Heart' cultivar.  
489 Nanoparticles can, therefore, serve as effective elicitors to boost metabolite production.

#### 490 ***Effect of plant growth regulators on the content of metabolites in callus***

491 The biochemical composition of callus depended on the type of explant from which it formed  
492 (Kulus and Tymoszuik 2020), consistent with reports from other authors (Hussain et al. 2012).  
493 In *L. spectabilis*, callus regenerating from petioles was the most abundant in carotenoids and  
494 anthocyanins, while callus from whole leaves contained the least pigments, suggesting a  
495 positive impact of stress (in this case, mechanical damage to the explant) on the biosynthesis  
496 of these compounds. The auxin-to-cytokinin ratio in the medium partially influenced the  
497 chemical profile of callus (i.e., chlorophyll and carotenoid content) but did not determine  
498 anthocyanin and total polyphenol content. Generally, callus formed on a medium with NAA  
499 contained the highest concentrations of the studied metabolites. This auxin can thus be used as  
500 an elicitor to control the overproduction of desired organic compounds in *L. spectabilis*  
501 culture. Interestingly, a negative correlation was observed between the biochemical  
502 compound content in the callus and the efficiency of somatic embryogenesis. Conversely, a  
503 high frequency of non-embryogenic callus formation was associated with higher  
504 concentrations of photosynthetic pigments, anthocyanins, and polyphenols (Kulus and  
505 Tymoszuik 2020). This suggests that the chemical composition can be used as a marker of the  
506 embryogenic potential of cells and *vice versa*. A positive correlation was also found between  
507 the content of all studied chemical groups in *L. spectabilis* callus, allowing for the

508 simultaneous overproduction of these substances. Comparative analyses, however, showed  
509 significant differences in pigment content between callus and shoots (Kulus 2020a).

#### 510 ***Enzymatic activity of plants in tissue culture systems***

511 The unnatural conditions in the *in vitro* culture can lead to the release of ROS, causing lipid  
512 peroxidation and disturbances in the redox state of cells, resulting in oxidative stress (Timoteo  
513 et al. 2019). Enzymatic antioxidant mechanisms catalyze the breakdown of ROS, and changes  
514 in enzyme activity are considered biological markers of oxidative stress (Homae and  
515 Ehsanpour 2016). Colorimetric and kinetic reaction studies allowed for understanding the  
516 enzymatic response of *L. spectabilis* cells subjected to various stressors (Kulus and Tymoszuk  
517 2021; Kulus et al. 2022). Both in cryopreservation (Kulus and Tymoszuk 2021) and induced  
518 mutagenesis studies (Kulus et al. 2022), the highest increase in activity was observed for  
519 guaiacol peroxidase (GPOX), while the lowest was for glutathione reductase (GR). Therefore,  
520 GPOX is considered the most sensitive marker of oxidative stress in *L. spectabilis*, being the  
521 most involved in the plant's defense mechanism. Despite long-term exposure to nanoparticles,  
522 this factor generally caused the least changes in enzyme activity compared to other, more  
523 short-term stressors (Kulus et al. 2022). This underscores the suitability of using AuNPs in *L.*  
524 *spectabilis in vitro* systems.

525

#### 526 **Somaclonal variation and induced mutagenesis in expanding the genetic variation in** 527 **bleeding heart**

528 Breeding programs conducted in the 20th and 21st centuries have led to the creation of new  
529 cultivars of bleeding heart, although their number is still limited (Hodges 2012). In Korea,  
530 effort is made to develop commercial cultivars of this species through transgenesis (Lee and  
531 Lee 2003). However, genetic engineering is a complex, time-consuming, and costly method  
532 of plant improvement, especially in the case of bleeding heart, for which the genome size was  
533 not even known until 2022. This method is also subject to public debate. The use of  
534 somaclonal variation induced by *in vitro* culture conditions can be a cheaper, though equally  
535 effective approach (Miler and Zalewska 2014; Duta-Cornescu et al. 2023).

536 In the study performed by Kulus et al. (2021), it was found that bleeding heart is  
537 susceptible to somaclonal variation induced by the simultaneous presence of auxins and  
538 cytokinins in the culture medium. Variation at the DNA sequence level was detected in 36.8%  
539 and 69.1% of plants by SCoT and RAPD markers, respectively. It was probably associated  
540 with the development of genetically unstable callus at the base of shoots or exposure of plants  
541 to intensively secreted polyphenols into the medium. Additional cluster analyses and



542 calculated genetic distance between individuals considered most of this variation  
543 insignificant; however, four newly obtained genotypes differed significantly from the control.  
544 Due to the lack of evident phenotypic changes, the use of more "aggressive" agents for  
545 breeding purposes was indicated (Kulus et al. 2021).

546 Classical mutation breeding, based on random induction of mutations, is an attractive  
547 alternative to costly and difficult genetic engineering procedures (Miler et al. 2021).  
548 Mutagenic factors used in plant breeding can be classified as chemical and physical. Among  
549 chemical mutagens, ethyl methanesulfonate (EMS) is most commonly used (Shelake et al.  
550 2019); however, recent studies have shown that nanoparticles can also be effectively used to  
551 induce genetic variability in plants due to their ease of absorption by cells and interaction with  
552 proteins and DNA (Mehrian and De Lima 2016; Tymoszuk and Kulus 2020). Physical  
553 mutagens, such as X-rays or gamma radiation, are more environmentally friendly than  
554 chemical mutagens, as their use does not generate harmful waste. Unfortunately, the  
555 availability of irradiation devices is a limiting factor for plant breeders, as these devices are  
556 usually owned by national atomic energy agencies or medical/scientific institutes (Miler et al.  
557 2021). A cheap source of electromagnetic radiation useful in inducing variability can also be a  
558 classic microwave oven. Microwaves have been used in mutation breeding of  
559 *Chrysanthemum × morifolium* (Ramat.) Hemsl. (Miler and Kulus 2018).

560 In the study by Kulus et al. (2022), the effect of gold nanoparticles (AuNPs),  
561 microwaves (non-ionizing radiation), and X-rays (ionizing radiation), applied in various  
562 doses, was investigated on plant acclimatization efficiency and the possibility of using these  
563 factors in mutation breeding and inducing variability at the genetic and phenotypic level in  
564 bleeding heart.

565 It is worth emphasizing the high survival rate of primary explants in the *in vitro* culture,  
566 which ranged from 86% to 100% (Kulus et al. 2022). This is significantly higher than in other  
567 species treated with mutagens (Tallón et al. 2015) and may indicate high resistance of *L.*  
568 *spectabilis* to this type of stress. Bleeding heart may therefore constitute a valuable source of  
569 genes in breeding programs based on crossing, hybridization, and transgenesis. It was found  
570 that plants from the untreated control and plants obtained after the longest period of  
571 microwave irradiation more often died during acclimatization than those from other  
572 experimental treatments. Control plants were also of noticeably poorer quality during further  
573 greenhouse cultivation (Kulus et al. 2022). The hormetic response may therefore be a strategy  
574 by which *L. spectabilis* acclimates to more difficult conditions, leading to speciation in the  
575 evolutionary process (Małkowski et al. 2020).

576 Cytometric analyses allowed to determine that *L. spectabilis* has a very small genome  
577 (1281 Mbp; 1.314 pg/2C DNA), according to the classification by Soltis et al. (2003), which  
578 makes it a good model species for experimental biology research, concerning, for example,  
579 directions of evolution. No changes in ploidy were detected among the studied plants, but in  
580 several individuals treated with X-rays, nanoparticles, or microwaves, significant changes in  
581 nuclear DNA content were found (Kulus et al. 2022). In most cases, there was a decrease in  
582 DNA content (most likely due to deletions), and in one case, an increase in the value of this  
583 parameter (which can be explained by the transition of somatic cells into endocycle). A  
584 similar phenomenon was observed, for example, in chrysanthemum (Miler et al. 2020). The  
585 greatest fluctuations in DNA content were found after applying the highest dose of X-rays,  
586 suggesting that this factor affects bleeding heart plants to the greatest extent.

587 The effectiveness of the applied mutagens in inducing variability in *L. spectabilis* was  
588 confirmed by SPAR (single primer amplification reaction) genetic analyses (Kulus et al.  
589 2022). This confirms the usefulness of this group of markers in poorly studied species. PCR-  
590 based genotyping methods are still widely used to assess plant genetic variability due to their  
591 simplicity and versatility (Nadeem et al. 2017). Mutations were detected in 7.5% of plants by  
592 DAMD (Directed Amplification of Minisatellite DNA (Heath et al. 1993)), RAPD, and SCoT  
593 markers, but not by ISSRs. Therefore, the discriminatory power of the studied molecular  
594 marker systems in *L. spectabilis* can be arranged in the following order:  
595 ISRR<SCoT<RAPD<DAMD (Kulus et al. 2022). The lack of species-specific primers  
596 confirmed the usefulness of primers previously used in other botanical families. Interestingly,  
597 most polymorphisms were detected by the DAMD primer 5'-  
598 AATGTGGGCAAGCTGGTGGT-3' (Seyedimoradi et al. 2012), although the PCR thermal  
599 profile for this marker system was the most difficult to optimize. Phenotypic changes  
600 concerning leaf shape were identified in plants irradiated with X-rays or, less frequently, in  
601 plants treated with AuNPs. Despite the observed changes in DNA sequence, no permanent  
602 phenotypic changes were found in objects treated with microwaves, suggesting that this is the  
603 least effective of the studied mutagens (Kulus et al. 2022). These experiments create  
604 perspectives leading to obtaining new cultivars of *L. spectabilis* plants with increased  
605 horticultural and/or phyto-pharmaceutical potential.

606

## 607 **Conclusions**

608 Research indicates that in bleeding heart, the activation of axillary buds *in vitro* (especially in  
609 the presence of KIN in the medium) and the induction of indirect somatic embryogenesis (in

610 the presence of PIC) are significantly easier to achieve than the regeneration of adventitious  
611 shoots. There are evident cultivar differences within the species. The 'Valentine' cultivar is the  
612 easiest to culture *in vitro*, while 'White Gold' is the most challenging. Since classical growth  
613 regulators can cause anatomical, physiological, and biochemical disturbances in bleeding  
614 heart, it is advisable to search for new substances stimulating morphogenesis *in vitro*.  
615 Examples of such substances may include nanoparticles or substances of natural origin. The  
616 negative effects of synthetic growth regulators can be avoided by supplementing the medium  
617 with coconut or rice extract (low in polyphenols and high in saturated fatty acids), depending  
618 on the preferences of individual cultivars. The addition of gold nanoparticles, on the other  
619 hand, positively affects shoot proliferation and the quality of the obtained plantlets, which are  
620 characterized by higher survival rates after transferring to *ex vitro* conditions. Simultaneous  
621 overproduction of bleeding heart metabolites can be achieved using exogenous auxins (IAA,  
622 NAA) and gold nanoparticles. Analysis of cellular enzymatic activity identified guaiacol  
623 peroxidase as the most sensitive marker of oxidative stress in *L. spectabilis* caused by water  
624 deficit or thermal factors. The most stable among the studied catalytic proteins under stress  
625 conditions is glutathione reductase. It was demonstrated that cryogenic techniques based on  
626 explant encapsulation are the most effective in terms of biological material viability, its  
627 further growth, and genetic stability. Nanomaterials in low concentrations can increase the  
628 effectiveness of cryogenic techniques by over 20%. This discovery may be groundbreaking  
629 for the further development of cryobiology, although the final effect depends also on their  
630 size. The addition of coconut extract to the medium also positively affects shoot proliferation  
631 after "thawing" of explants. On the other hand, sesame extract can be used as a natural  
632 retardant, for example, in slow-growth plant culture. The effectiveness of tested mutagens in  
633 mutation breeding of bleeding heart can be ranked as follows: microwaves < AuNPs < X-  
634 rays. Although individual SPAR marker systems have certain limitations, the collective data  
635 obtained from several types of systems provide a comprehensive description of genetic  
636 variability within the species.

637

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640

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649

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958 **Fig. 1.** Four commercial cultivars of *L. spectabilis*: ‘Alba’ (A), ‘Gold Heart’ (B), ‘White  
959 Gold’ (C), and the most recent one ‘Valentine’ (D) cultivated at the Bydgoszcz University of  
960 Science and Technology, Poland.

961

962 **Fig. 2.** *In vitro* growth of single-node explants of *L. spectabilis* on the MS medium devoid of  
963 plant growth regulators (A), the presence of auxins and cytokinins (PGRs) stimulates the  
964 secretion of harmful phenolics into the medium (B), gold nanoparticles (AuNPs) in the culture  
965 medium stimulate the most abundant growth of good-quality shoots (C), non-meristematic  
966 explants (leaves and internodes) regenerate only embryogenic callus (D), complete  
967 micropropagated plants grown *ex vitro*.

968

969 **Fig. 3.** Comparison of the effectiveness of various cryopreservation protocols in *L.*  
970 *spectabilis*: V – vitrification, D-V – droplet-vitrification, E-V – encapsulation-vitrification, E-  
971 D – encapsulation-dehydration.

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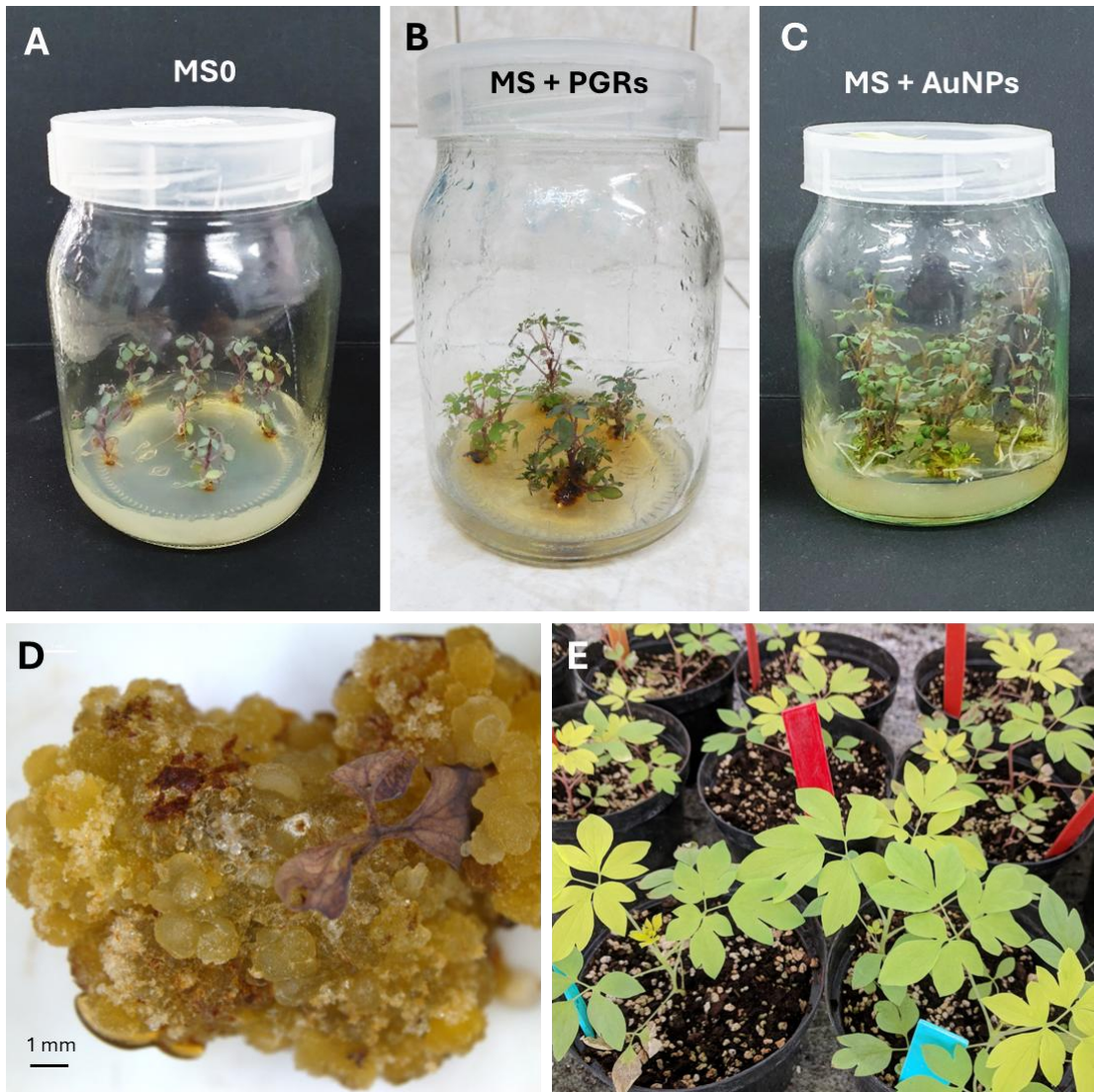
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977 Fig. 1



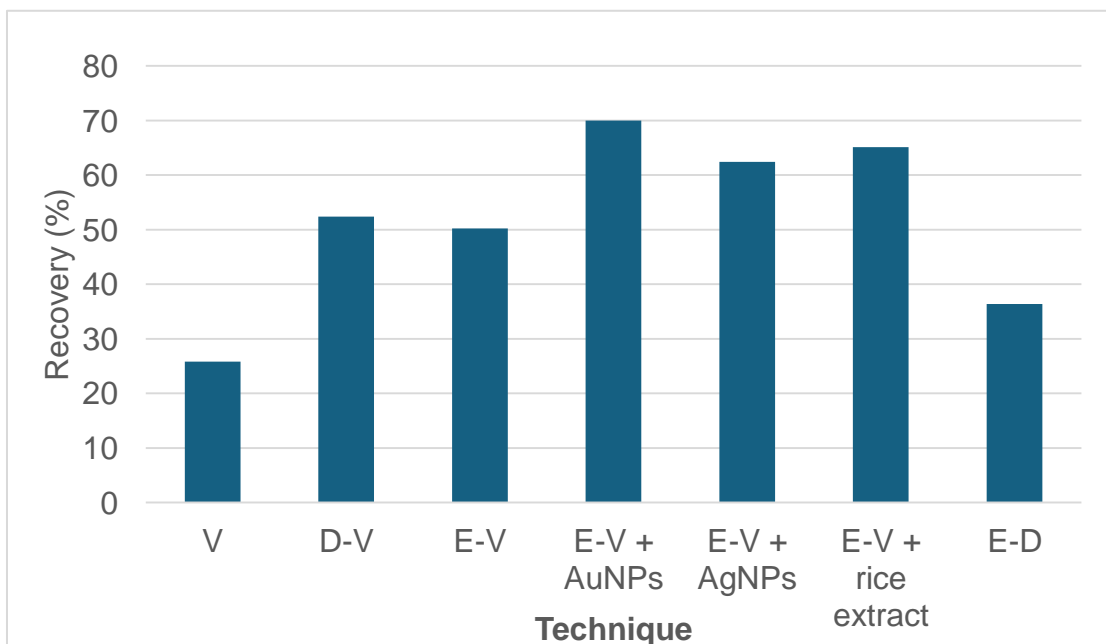


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979 Fig. 2

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1001 Fig. 3